



Micro-extraction of Aqueous 1,4-Dioxane Samples for Analysis by GC/MS SIM

Analytical Procedure for 1,4-dioxane
extraction from 200 μL of water with a
20- $\mu\text{g}/\text{L}$ reporting limit

Background and Assignment

- Test samples are bioremediation small-scale tests that must be sampled over time without significant change to the test system.
- Goal was to get below 50 ppb and to go as low as possible.
- Total available sample volume per test limited to 200 μL .
- Personnel time, equipment, and funding were limited.
- Target compound 1,4-dioxane has both very poor purging efficiency and poor extraction efficiency due to its high solubility.



Eliminations

- Testing via direct aqueous injection (DAI) sensitive to only 500 µg/L.
- Purge and Trap just not sensitive enough
- Conventional liquid-liquid extraction would require time consuming extraction and would require sample volumes beyond what was available.

Project Planning Options

Plan A:

- Frozen micro extraction.
- Take 200 μL of sample and add 200 μL of methylene chloride in a 2 mL instrument vial.
- Freeze sample.
- Syringe off methylene chloride
- Dry and analyze

Plan B:

- Adapt Solid Phase Micro Extraction (SPME) for 200 μL sample.
- Manual extraction set up on rotatory shaker for agitation.
- Direct Manual injection



Pros and Cons

Frozen Micro Extraction (Plan A)

- Very low on time and materials
- Unproven technique
- Requires GC/MS SIM

SPME (Plan B)

- Potentially a sensitivity multiplier
- Generally proven technology, but untested for volume/scale limitations of project
- Typically requires autosampler requiring higher volume for high precision
- 200- μ L extraction would requires manual set-up

Method Selection Considerations

- Micro extraction (Plan A) was selected because it appeared to be relatively simple to try and much of the equipment was readily available
- Equipment was purchased for manual SPME (Plan B) and experimental design was developed as a backup if Plan A did not work.

Final Experimental Design for Frozen Micro-extraction

- 200- μ L aliquot of sample(s) is transferred to a screw top auto-sampler vial(s).
- Sample(s) is spiked with 200 ppb of 1,4-dioxane- d_8 and 100 ppb of 1,4-dichlorobenzene- d_4 .
- Vials are placed in a -80°C freezer at a 45° angle for 30 minutes.
- Vials are taken out of freezer individually and are immediately prepared for analysis.

Experimental Design for Frozen Micro-extraction (Cont.)

- Methylene chloride solvent is syringed from the vial and transferred to a clean instrument vial fitted with a 200- μ L insert.
- Sodium sulfate is added to vial to fill the bottom tip of the insert approximately 1/16 of an inch.
- Vials may be placed directly on an instrument for analysis.
- A 5975 instrument was set up in GC/MS SIM mode for analysis.

A banner image showing a sunset over a body of water with silhouettes of trees and animals in the foreground. The sky is filled with orange and yellow clouds, and the sun is low on the horizon.

Analytical Methods

- Holding Time Considerations
- Instrumentation and Equipment
- Instrument Program Settings
- Quality Control & Acceptance Criteria

Holding Time Considerations

Prep	Method	Matrix	Preservation	Holding Times
Frozen – micro extraction	8270C SIM	Aqueous	Cool to 4°C	ASAP
SPME	8270C 8260B	Aqueous	Cool to 4°C	48 hours

- Holding times were shortened for Method 8270C due to risk of biodegradation of the 1,4-dioxane in study.
- Holding time for SPME represents time from extraction to injection.



GC/MS Instrument

Sample Introduction	Agilent 7683B AutoSampler
Instrument	Agilent 5975 GC/MS
GC Column	HP-5, 30 m x 250 μm x 0.25 μm
Injection Volume	1 μL
Solvent	Methylene Chloride
EM Offset	200



GC Oven Parameters

- 30°C Hold for 5 minutes
- Ramp 15°C/minute to 100°C
- Hold for 0 minute
- Ramp 50°C/minute to 275°C
- Hold for 1 minute

SIM Acquisition Parameters

- Solvent Delay 5.00 minutes
- SIM Group 1 Start Time 5.00
- Group 1 - (Mass, Dwell)
(58.00, 100) (88.00, 100) (96.00, 100)
- SIM Group 2 Start Time 8.00 minutes
Group 2 - (Mass, Dwell)
(150.00, 100)



GC/MS Instrument Tune

- 1,4-dioxane is a fairly low mass compared to many semi-volatiles.
- Recommended using BFB for the tune check
- SW-846 volatile criteria used for tune acceptance every 12 hours.
- When necessary PFTBA was used for autotune on the LOMASS program

A sunset over a body of water with silhouettes of trees and birds in the sky. The sun is low on the horizon, casting a warm glow across the sky and water.

Initial Calibration

- Established initially with a minimum of six concentration levels with low standard at or below limit of quantitation (LOQ).
- Calibration blank is included in the curve.
- Both the initial calibration (ICAL) and the continuing calibration verification (CCV) are prepared in water and extracted in the same manner as the samples.

Initial Calibration (Cont.)

- When the percent relative standard deviation (%RSD) for target compound is $\leq 15\%$, an average response factor may be used.
- When %RSD for target compound is $>15\%$, a linear regression or quadratic curve must be used.
- A regression curve must have a correlation coefficient (R^2) ≥ 0.99 .
- Force through zero was turned off.

Continuing Calibration Verification

- CCV is run every 12 hours immediately after the instrument tune.
- Should be at or near the mid-point calibration range for the target compound.
- Minimum %D for CCV should be $\leq 30\%$ (recommended).
- When possible, the CCV should be prepared using a separate dilution series from neat to verify primary calibration accuracy.



Method Blank

- One Di Water and reagent blank per batch of up to 20 samples per day; run with samples.
- Must undergo ***all*** sample preparative and cleanup procedures.
- All target compounds of interest must be \leq LOQ.

Laboratory Control Sample

- Two per extraction batch per set of 20 samples per day.
- The CCV and laboratory control sample (LCS) are equivalent for this method.
- The CCV/LCS were prepared as a separate working standard from neat material.
- The default recovery recommended for this method is $100 \pm 30\%$.
- RPD is considered acceptable if the successive LCSs are within range.

A banner image showing a sunset over a body of water with silhouettes of trees and birds in the sky. The text 'Sample Replicate' is overlaid in white on a dark blue background.

Sample Replicate

- One set per extraction batch per set of 20 samples per day.
- The default % RPD = 30%.
- Alternatively, project control limits may be calculated using $\pm 3 \times$ the standard deviation of recoveries.

Qualitative/Quantitative Issues

- The raw result for 1,4-dioxane must be within the calibration range (\leq highest standard) for all samples.
- Compounds exceeding the instrument calibration range (highest calibration standard) must be diluted to within calibration range and reanalyzed.
- Analyst should take precautions against carryover, including reanalysis of samples when carryover is suspected.



Qualitative/Quantitative Issues (Cont.)

- The blank analyzed immediately after any standard must display a concentration of the 1,4-dioxane $<$ the LOQ.
- Any unusual change in instrument chromatography should be investigated as a potential change in instrument performance.



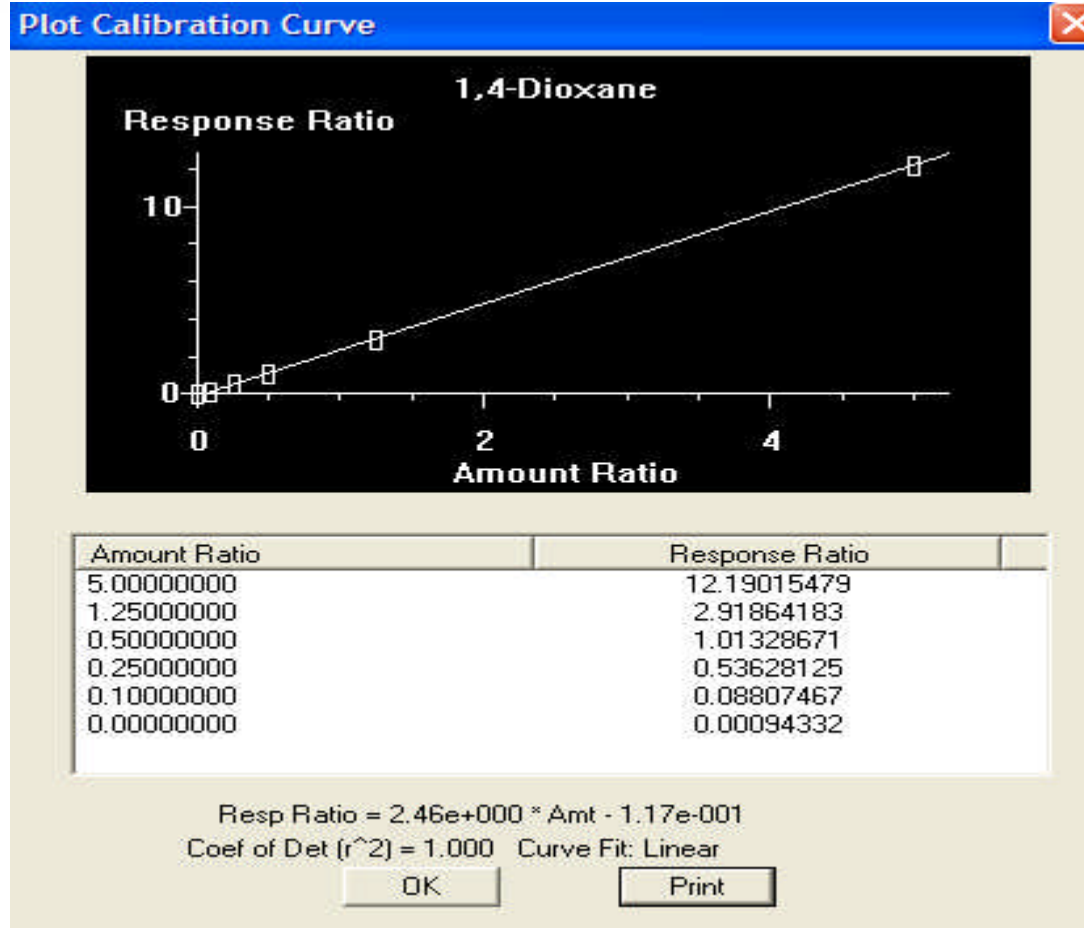
Method Performance Data

The following were put together for a final demonstration of method performance before writing the SOP.

- Curve Linearity and Range demonstration.
- Initial Demonstration of Capability Study Using LCS level spike.
- Low Level Demonstration of Capability as an LOQ verification.

Calibration Curve

1000 ppb
250 ppb
100 ppb
50 ppb
25 ppb
0 ppb



Initial Demonstration of Capability

Parameter	Spiked Amt (µg/L)	Average Recovery	Acceptance Range	STD DEV (s)	RSD
1,4-Dioxane	100.00	114.98%	70% - 130%	6.21 µg/L	5.40%
1,4-DCB-d ₄	200.00	111.99%	70% - 130%	13.06 µg/L	5.83%
1,4-Dioxane (Low level)	20.00	93.38%	70% - 130%	2.87 µg/L	15.36%



In Summary

- We developed a method for preparation and analysis of 200- μ L aliquot samples down to 20 μ g/L for 1,4-dioxane.
- All materials used were commonly available laboratory materials.
- Very reasonable time & costs.
- Acceptable linearity and accuracy to the reporting level.
- Detected values appear to be reliable to $\sim 1/2$ the RL or 10 ppb.

Thank You



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Patrick A. Conlon
Environmental Standards, Inc.
1140 Valley Forge Road
P.O. Box 810
Valley Forge, PA 19482
610.935.5577
solutions@envstd.com
www.envstd.com