Evaluation of Preservation Recommendations in SW-846 for Volatile Organic Chemicals in Aqueous Sample Matrices

Troy Strock, US EPA Region 5 Laboratory Shen-Yi Yang, US EPA Office of Resource Conservation and Recovery

All about me

- 2004 to present –US EPA Region 5 laboratory
 - GC/MS
 - Test various media for SVOCs and VOCs
- 2011-2012: 4 month detail to US EPA's Office of Resource Conservation and Recovery in OSWER
 - Worked with SW-846 methods team, Shen-Yi Yang and Kim Kirkland
 - Update V to 3rd edition of SW-846





Primary Goal of Preserving VOC Samples: Maintain Sample Integrity

Degradation Mechanisms:

- Biological
- Chemical (oxidation/reduction, hydrolysis)

Recommended sample preservation techniques:

- Store in low temperature environment ($\leq \&C$)
- Add pH modifier

SW-846, 3rd ed., Update IV (2007)

TABLE 4-1

RECOMMENDED SAMPLE CONTAINERS, PRESERVATION TECHNIQUES, AND HOLDING TIMES^a (Note: Footnotes are located on the last page of the table.)

VOLATILE ORGANICS							
	Sample Matrix	Preservative ¹	Holding Time ¹				
	Aqueous samples with no residual chlorine present	Cool to \leq 6 °C and adjust pH to less than 2 with H_2SO_4 , HCI, or solid NaHSO ₄	14 days				
		If carbonaceous materials are present, or if MTBE and other fuel oxygenate ethers are present and a high temperature sample preparative method is to be used, do not acid preserve the samples.	7 days				
		If vinyl chloride, styrene, or 2-chloroethyl vinyl ether are analytes of interest, collect a second set of samples without acid preservatives and analyze as soon as possible.	7 days				
	Solid samples (e.g. soils, sediments, sludges, ash)	See the individual methods.	14 days				
		If vinyl chloride, styrene, or 2-chloroethyl vinyl ether are analytes of interest, collect a second set of samples without acid preservatives and analyze as soon as possible.	7 days				

Holding Time / Preservation Table in SW-846 Method 5035A Appendix (2002)

Table A.1

Recommended VOC Sample Preservation Techniques and Holding Times

Sample Matrix	Preservative	Holding Time	Comment
Aqueous Samples With No Residual Chlorine Present	Cool to 4 ± 2°C.	7 days	If MTBE and other fuel oxygenate ethers are present and a high temperature sample preparative method is to be used, do not acid preserve the samples. If aromatic and biologically active compounds are analytes of interest, acid preservation is necessary and the holding time is extended to 14 days.
Aqueous Samples With No Residual Chlorine Present	Cool to 4 ± 2°C and adjust pH to less than 2 with HCl or solid NaHSO ₄ .	14 days ¹	Reactive compounds such as 2-chloroethylvinyl ether readily break down under acidic conditions. If these types of compounds are analytes of interest, collect a second set of samples without acid preservatives and analyze as soon as possible.

"PUBLIC COMMENT SUMMARIES AND RESPONSES UPDATE IV TO SW-846"

"Two commenters disagreed with the recommendation in Table 4-1 to collect a second set of samples without acid preservation if vinyl chloride is an analyte of interest. The commenters noted that some studies demonstrate that vinyl chloride is stable for at least 28 days and possibly over 90 days in acidic solution."

Response: The Agency agrees that vinyl chloride could be assigned a "highly variable stability" classification ...(depending on matrix conditions)... [I]t is also well known that reactive compounds; such as styrene, vinyl chloride, and 2-chloroethyl vinyl ether readily degrade under acidic conditions. For this reason, the current recommendation ...is that acid preservation should not be used if these compounds are target analytes.

Question to answer for Update V:

Does acid preservation during sampling create more problems for quantitative analysis of styrene and vinyl chloride than lack of preservation, or more problems than for other VOCs?

Sources of information:
1) Methods from other EPA programs
2) Literature search for VOC holding time studies
3) Wastewater holding time studies

CLP Guidance for Field Samplers, 2011:

Soils: "NaHSO₄ preservation creates low pH conditions that will cause the destruction of certain CLP target analytes (e.g., vinyl chloride, trichloroethene, trichlorofluoromethane, cisand trans-1,3-dichloropropene)." But no similar problems noted for aqueous matrices.

Drinking water method 524.3:

Styrene and vinyl chloride are target analytes, but no indication that maleic or ascorbic acid preservatives cause stability problems

Wastewater method 624:

• "Experimental evidence indicates that some aromatic compounds, notably benzene, toluene, and ethyl benzene are susceptible to rapid biodegradation under certain environmental conditions. Refrigeration alone may not be adequate to preserve these compounds in wastewaters for more than seven days. For this reason, a separate sample should be collected, acidified, and analyzed when these aromatics are to be determined."

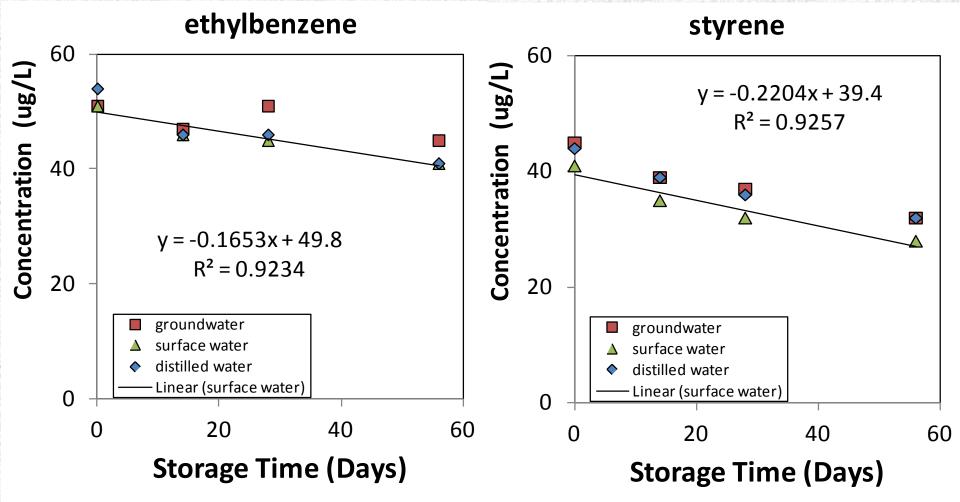
Vinyl chloride listed as a target analyte, but not styrene

Refrigerated holding time studies

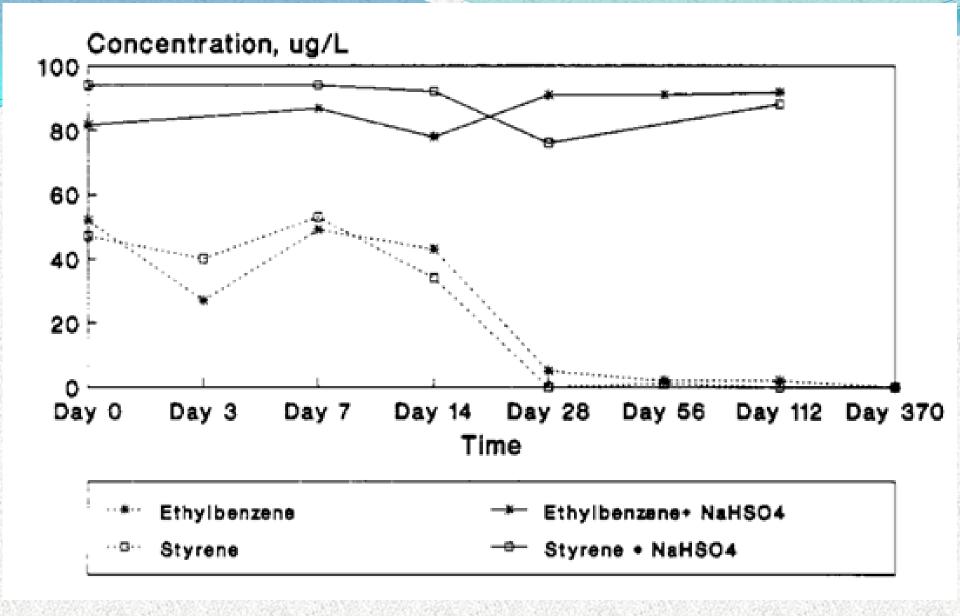
- Department of Energy / Oak Ridge National Lab
 - -Maskarinec et al (1989), unpreserved or preserved with bisulfate, HCl
 - West et al (1996), preserved with bisulfate
- USGS (1999), surface water and groundwater preserved with HCl
- Groundwater US EPA ORD Lab in Ada
 - preserved with HCl or trisodium phosphate
- Wastewater: Hampton Roads Sanitation District (VA)
 - Municipal and Industrial WWTP influent, unpreserved or preserved with HCl
- Wastewater Test America
 - Hydrofracking wastewater unpreserved or preserved with HCl

HCl preserved DI water, surface

water, and groundwater

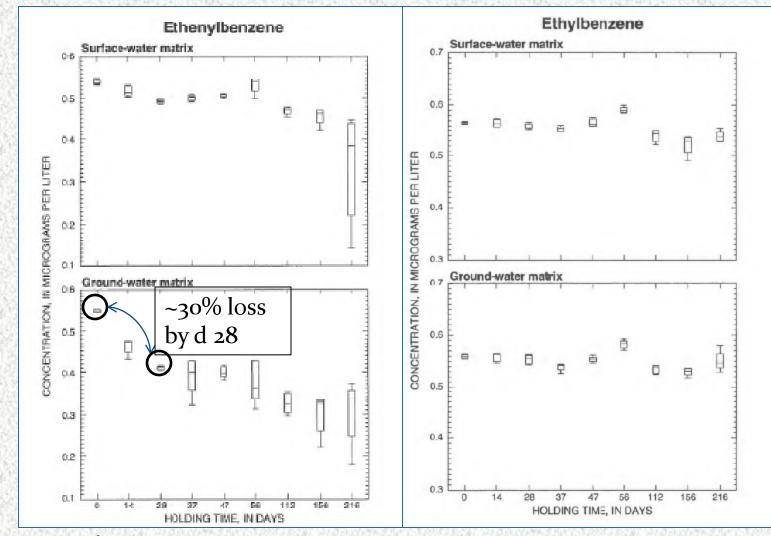


Maskarinec et al., 1989



Maskarinec et al., 1989

216 Day Surface and Groundwater Holding Time Study – Styrene



Love, et al., 1999.

Styrene Stability during 28 days

refrigerated storage

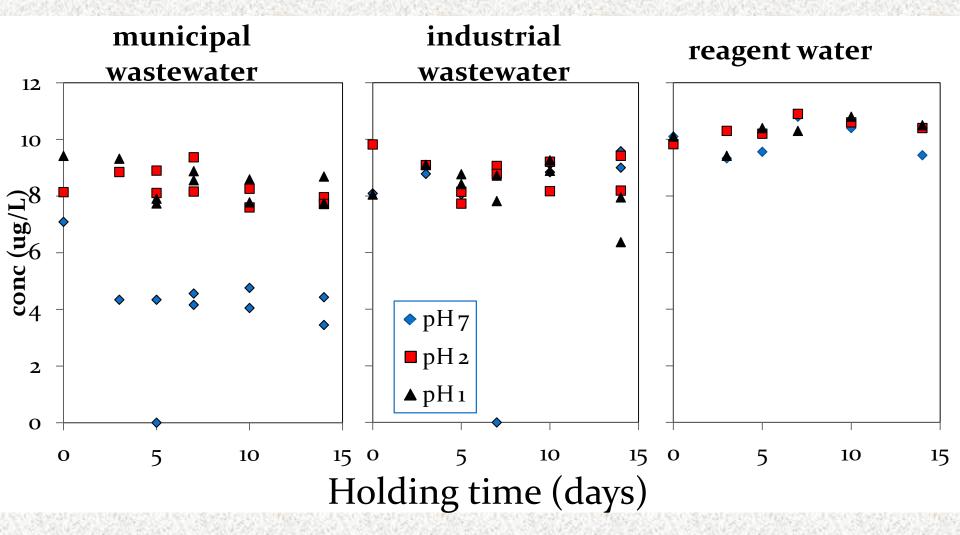
Groundwater from hazardous waste site investigation

Matrix spike recovery in pH 2 HCl preserved groundwater samples at d o and d 28

Spike level (ug/L)	d 0 mean % recovery	d 28 mean % recovery	S _{pool} (%)	% diff (d28 - d0)	Significant loss? (α=0.05)
20	93	69	6	24	Y
200	92	79	12	13	N

Data adapted from: Wilson, Fine, and Beach, 2008.

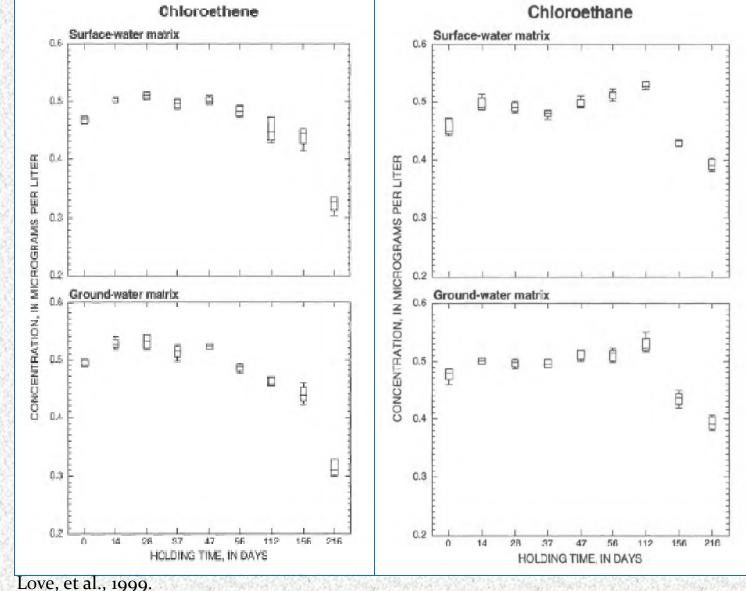
Municipal and Industrial Wastewaters Holding Time Study: Styrene



Data provided by Hampton Roads Sanitation District

216 Day Surface and Groundwater

Holding Time Study – Vinyl Chloride



Vinyl Chloride Stability during 28 days storage

of acid or base preserved samples

Matrix spike recovery in pH 2 HCl preserved groundwater samples at d o and d 28

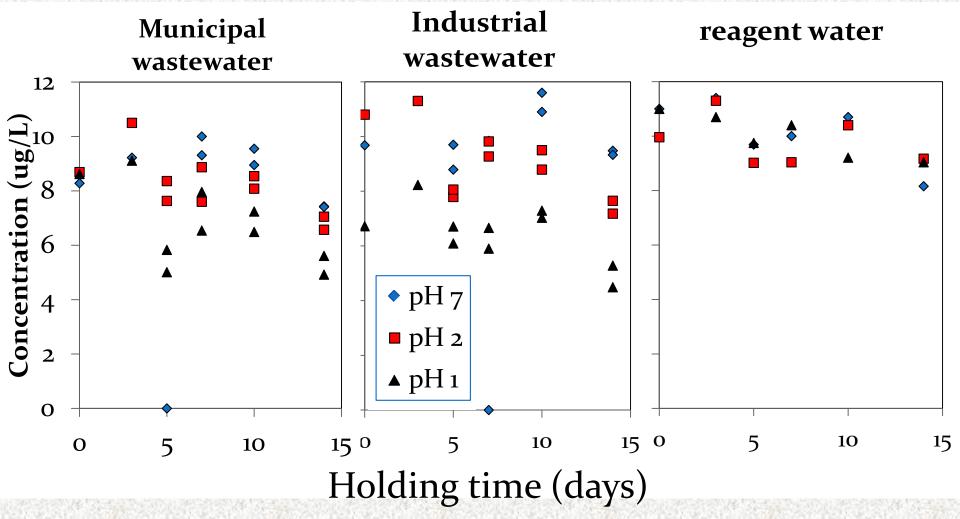
-	d 0 mean %			% diff	Significant loss?
(ug/L)	recovery	% recovery	S _{pool} (%)	(d28 - d0)	(α=0.05)
20	81	71	16	10	Ν
200	89	74	7	15	Y

Measured concentrations of select VOCs in acid or base preserved replicate samples after 28 days refrigerated storage.

Chemical	Mean sample conc (ug/L)	preservative	conc ratio (VC / cisDCE)	conc ratio in different preservatives (HCl / TSP)
	7480	HCI	0.92	0.94
vinyl chloride	7980	TSP	0.95	
cis-1,2-DCE	8170	HCI		0.97
	8400	TSP		· · · · · · · · · · · · · · · · · · ·

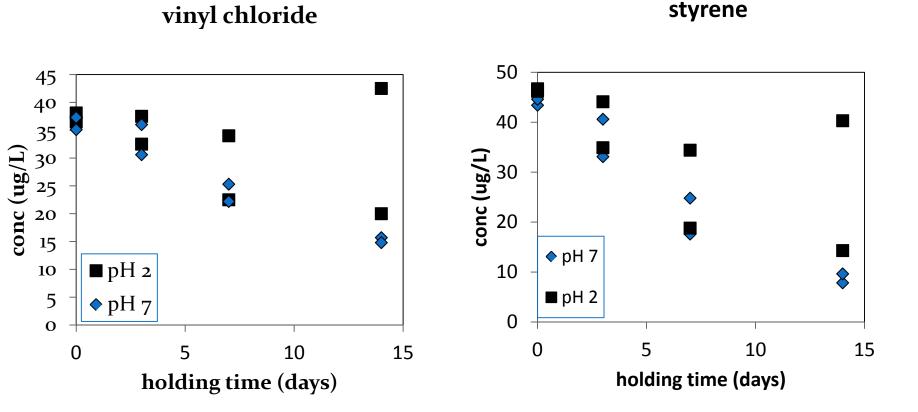
TSP – Trisodium Phosphate, HCl – Hydrochloric Acid Data adapted from: Wilson, Fine, and Beach, 2008.

Municipal and Industrial Wastewaters Holding Time Study: Vinyl Chloride

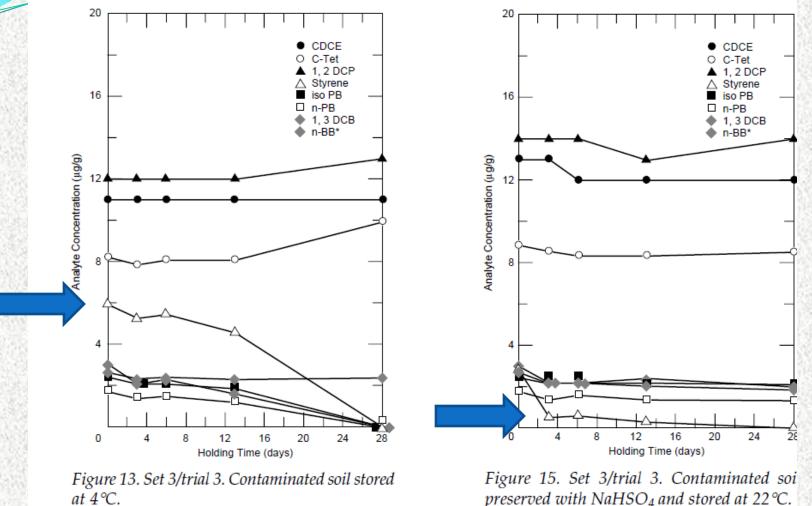


Data provided by Hampton Roads Sanitation District

Hydraulic fracturing wastewater holding time study from Test America



Stability in NaHSO4 preserved soil



"Styrene was not stable in the soil preserved with NaHSO4... The chemical reaction that transforms styrene most likely is catalyzed by the soil. Clearly, soil sample preservation by NaHSO4, or perhaps any acid, would not be compatible for investigations where styrene is a constituent of interest." A. Hewitt, 1995.

Observations from Study Data

• Styrene

- Some loss observed over extended holding times in acidified waters, but generally ≤ 30% over 28 days
- Significant losses of styrene in some unpreserved samples
- Vinyl chloride
 - Similar stability seen with acidic and basic preservative
 - No rapid losses resulting from acid preservation in sample matrices tested

Revised Table 4-1 in Update V

TABLE 4-1 (continued)

RECOMMENDED SAMPLE CONTAINERS, PRESERVATION TECHNIQUES, AND HOLDING TIMES*

VOLATILE ORGANICS (continued)				
Sample Matrix	Container ¹	Preservative ²	Holding Time ^s	
Aqueous samples WITH residual chlorine present	Methods 5021, 5030, 5031, and 5032: 2 x 40-mL vials with PTFE-lined septum caps	Collect sample in a 125-mL container which has been pre-preserved with 4 drops of 10% sodium thics little solution. Gently swirl to mix sample and transfer to a 40-mL VOA vial. Cool to ± 5 °C and adjust pH to less than 2 with H ₂ SO ₆ , HCI, or solid NaHSO ₆ .	14 days	
		If carbonaceous materials are present, or if MTBE and other fuel oxygenate ethers are present and a high temperature sample preparative method is to be used, do not acid preserve the samples.	7 days	
		If compounds that readily degrade in acidified water (e.g., 2-chloroethyl vinyl ethers) are analytes of interest, collect a second set of samples without acid preservatives and analyze as soon as possible.	7 days	
Acrolein and Acrolonicile	Methods 5021, 5030, 5031, and 5032:	Adjust to pH 4-5. Cool to 35 °C.		
Aqueous samples	2 x 40-ml vials with PTFE-lined septum caps	These compounds are highly reactive and should be analyzed as soon as possible.	7 days	
Solid samples (e.g. solis, sediments, sludges, ash)	Method 5035: See the method. Method 5021: See the method. Methods 5031 and 5032: See the methods.	See the Individual methods. If compounds that may be reactive in acidified solis (e.g., vinyl chloride, styrene, 2-chloroethyl	14 days	
		vinyl ether) are analytes of interest, collect a second set of samples without acid preservatives and analyze as soon as possible.	7 days	

A word about preservation

"If a preservative functions as intended, the concentrations of the analyte of concern will not change during storage. Science can not be used to prove a negative. Experimental trials with preservatives can not be used to prove that a preservative is universally effective."

- John T. Wilson, microbiologist at EPA ORD NRMRL, in "A Guide for Assessing Biodegradation and Source Identification of Organic Ground Water Contaminants using Compound Specific Isotope Analysis", EPA 600/R-08/148, 2008

Thanks

Workgroup

- Chemists from USEPA Regions 3, 4, 6, 10
- Jessie DeLuna, Paula Hogg, HRSD
- Richard Burrows, Test America
- Chung-Rei Mao, US ACE
- John Wilson USEPA ORD NRMRL

References

- Maskarinec, Bayne, Johnson, Holladay and Jenkins. Stability of Volatile Organic Compounds in Environmental Water Samples: Storage and Preservation. 1989. Oak Ridge National Laboratories Publication # ORNL TM-11330.
- A. D. Hewitt. Enhanced Preservation of Volatile Organic Compounds in Soil With Sodium Bisulfate. US ACE CRREL. Special Report 95-26. 1995.
- West, Bayne, Siegrist, Holden, Scarborough, and Bottrell. Stability of Purgeable VOCs in water samples during Pre-analytical holding: Part 1. Analyses by a Commercial Laboratory; and, Part 2. Analyses by an EPA Regional Laboratory. Oak Ridge National Laboratory. ORNL/TM-13240/V1 (1996) and ORNL/TM-13240/V2 (1997).
- A. B. Dindal, R. A. Jenkins, C. K. Bayne, D. W. Bottrell, D. Carden. Evaluation of the Impact of Post-Holding Time Analyte Degradation on Regulatory Decision-Making VOCs in Water. 2001.
- J. T. Love, G. C. Delzer, S. R. Abney, and J. S. Zogorski.Study Design and Analytical Results Used to Evaluate Stability of Volatile Organic Compounds in Water Matrices. USGS. 1999.
- J. T. Wilson, D. D. Fine, and S. Beach. Effects of Acid and Base Preservation on the Recovery of 48 VOCs During Storage and Analysis. In: Guide for Assessing Biodegradation and Source Identification of Organic Ground Water Contaminants using Compound Specific Isotope Analysis. 2008.